CHEMICAL PATHOLOGY TESTS REQUESTING & SPECIMEN COLLECTION

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INTRODUCTION

- Chemical Pathology deals with analyzing a wide range of substances (e.g. electrolytes, blood gases, enzymes, glucose, proteins, lipids, hormones, etc.) in the blood and body fluids.
- ~70% of clinical decisions are made based on laboratory tests.¹
- The result of any laboratory test is only as good as the specimens received in the laboratory.

¹Rohr UP, Binder C, Dieterle T, Giusti F, Messina CGM, Toerien E, et al. The value of in vitro diagnostic testing in medical practice: a status report. PLoSOne 2016;11(3): e0149856.

EFFECTS OF UNSUITABLE SPECIMENS

- Compromise the validity of test results
 → misdiagnosis, improper treatment,
 malpractice.
- Requires a repeat collection from the patient ? Precious sample.
- Delay in diagnosis/ treatment/ patient care.
- Patient discomfort and dissatisfaction.
- Additional expenses.



FREQUENCY & TYPE OF LABORATORY ERRORS

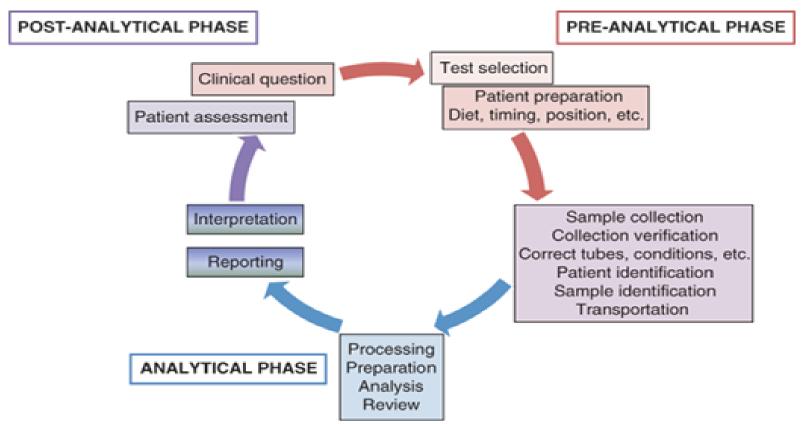
	Absolute frequency (ppm)		Relative f (%)	requency
	1996	2006	1996	2006
Total errors	4667	3092		
Preanalytic	3186	1913	68.2	61.9
Analytic	617	646	13.3	15.0
Postanalytic	864	715	18.5	23.1

Plebani M. The detection and prevention of errors in laboratory medicine. Annals of Clinical Biochemistry. 2010;47(2):101-110.

preanalytical error priːænəˈlɪtɪkəl ˈɛrə

noun

Preanalytical errors are errors in test results that occur as a consequence of actions or events preceding the test or analysis itself.



Source: Howard M. Reisner: Pathology: A Modern Case Study www.accessmedicine.com Copyright © McGraw-Hill Education. All rights reserved.

SPECIMEN REJECTION CRITERIA

- Defective / Missing / Wrong label
- Incomplete Request Form
- No specimen received (only request form received)
- Haemolysed / clotted sample
- Lipaemic / Icteric sample
- Insufficient specimen
- Wrong / Expired / Broken / Leaking collection containers

- Improper Specimen Collection (e.g. Wrong order of draw)
- Repetitive test requests

 (e.g. HbA1c request is < 8 weeks from previous testing)
- Improper transportation method / temperature not maintained
- Out of sample stability / Delayed sample arrival
- Test is not clinically indicated

 (e.g. Free PSA is rejected when total PSA result is not within 2.5 10 ng/ml)
- Test is not offered

Clinical Laboratory Handbook 1st edition, page 5. Downloadable from <u>https://pathologyuitm.weebly.com/</u>

CHEMICAL PATHOLOGY SPECIMEN REJECTION (JAN – JUNE 2020)

Rejection rate (SG BULOH)	506 out of 25,751 specimens (1.96%) <i>Target: < 1%</i>		
Top 3 Reasons	No. 1	No. 2	No. 3
for Rejection	REPETITIVE REQUEST	HAEMOLYSIS	CLOTTED SAMPLE
Percentage of rejection	35.1%	24.1%	19.2%





CHEMICAL PATHOLOGY SPECIMEN REJECTION (JAN – JUNE 2020)

Rejection rate (SELAYANG)	42 out of 13,465 specimens (0.31%)		
Top 3 Reasons	No. 1	No. 2	No. 3
for Rejection	NO SPECIMEN RECEIVED	REPETITVE REQUEST	HAEMOLYSIS
Percentage of rejection	52.4%	21.4%	11.9%





REPETITIVE REQUESTS

- Waste of resources

 (e.g. materials, reagents, lab staff time)
- Increases laboratory's workload
- Delay in turnaround time of other genuinely necessary tests
- Increases the rate of specimen rejection
- Increase in costs
- Patient discomfort and dissatisfaction

COMMONLY AFFECTED TESTS:

- HbA1c requested < 8 weeks from the previous testing</p>
- Lipid profile (FSL) requested repetitively during out of office hours.



HbA1c requests < 8 weeks from previous testing

- HbA1c represents the average rate of glycation over the lifespan of HbA (120 days), particularly in the previous 6-8 weeks.
- Glycation is a SLOW, irreversible process.

HbA _{1C} monitoring of patients with type 2 diabetes	2–6 monthly intervals (tailored to individual needs), until the blood glucose concentration is stable on unchanging therapy; use a measurement made at an interval of less than 3 months as an indicator of direction of change,	The Royal College of Pathologists Pathology: the science behind the cure Better Science. Better Testing. Better Care		
	rather than as a new steady state	National minimum retesting intervals in pathology		
	Six monthly intervals once the blood glucose concentration and blood glucose lowering therapy are stable	A final report detailing consensus recommendations for minimum retesting intervals for use in pathology		

Reminders in UniMEDS for HbA1c requests

For HBA1C kindly plea	se send to CPDRL within 2-4 hou	ITEST WILL BE REJECTED) IF REQUESTED WITHI	N 8 WEEKS Any special requ	est must be discussed with
Chemical Pathologist	on duty.				
					OK

Patholo	ogy Ord	er Conf	irmati	ion
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No	Test Name	Price(RM)	Form	Specimen	Test Location	Priority	Action
1.	BLOOD GASES (VENOUS)	32.00	- N/A -	BLOOD	CPDRL SG BULOH		DELETE
2.	HBA1C - Previous Sample Taken : 10-10-2017	16.50	- N/A -	BLOOD	CPDRL SG BULOH		DELETE
3.	OGIT	7.00	- N/A -	PLASMA	CPDRL SG BULOH		DELETE

Specimen collection : O Today

ay O Appointment

No.	Test Name	Special Instruction	Volume Required	Offsite test
1	HBA1C	Send to CPDRL within 2-4 hours.TEST WILL BE REJECTED IF REQUESTED WITHIN 8 WEEKS AFTER PREVIOUS REQUEST. Any special request must be discussed with Chemical Pathologist on duty.	3 ml	
2	RI OOD GASES (VENOUS)	Send to CDDRI immediately in ice	1 ml	

List of 24-hour (on-call) tests

- Blood Gases
- hs Troponin T
- Renal Profile
- Liver Function Test
- Bone Profile
- Glucose
- Calcium
- Magnesium
- Phosphate
- Bilirubin

- Creatine Kinase
- Amylase
- AST
- CRP
- Urine FEME (dipstick only)
- Urine Pregnancy Test
- Body Fluids
 Biochemistry

FSL is not part of the 24-hour / On-call tests. Specimens received out of office hours will be kept and analyzed on the next working day.

Clinical Laboratory Handbook 1st edition, page 11. Downloadable from <u>https://pathologyuitm.weebly.com/</u>

HAEMOLYSIS



Effects of haemolysis:

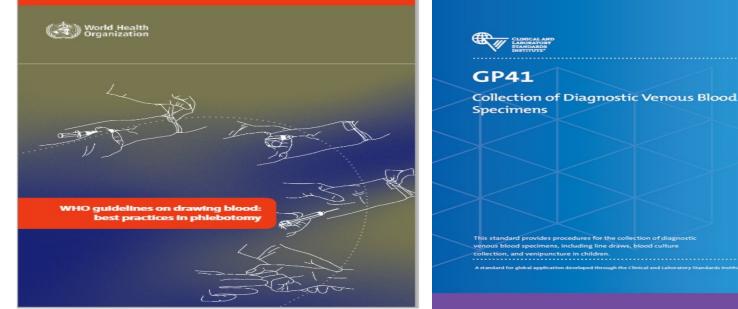
Hemolysis

- \circ Overestimation of **K**, ALT, AST, creatinine, CK, iron, LDH, lipase, Mg, PO₄, urea.
- Significantly decreased Na, albumin, ALP, Cl, GGT and glucose.
- \circ A hemolysis index of ~150 (Hb 1.9 g/L) caused > 20% change in cardiac **troponins** (the direction of change depends on assay).
- Clinically significant variations of AST, Cl, LDH, K and Na were observed in specimens displaying mild or almost undetectable hemolysis by visual inspection (Hb < 0.6 g/L).

Lippi G et al. Influence of hemolysis on routine clinical chemistry testing. Clin Chem Lab Med 2006;44(3):311–316 Renze Bais, The Effect of Sample Hemolysis on Cardiac Troponin I and T Assays, Clinical Chemistry 2010; 56(8), 1357–1359

PREVENTION OF HAEMOLYSIS

- Use suitable needle gauge (20-22 G for routine collection). Too small a needle results in excess vacuum force, too large a needle cause shear stress on the cell walls.
- Draw sample from antecubital region of the arm. Drawing from other sites has been shown to result in a higher degree of hemolysis.
- Warm up the puncture site to increase blood flow and prevents the need to "milk" the site.
- Do not leave the tourniquet on for > 1 minute. Prolonged tourniquet time causes the interstitial fluid to leak into the tissue, promoting hemolysis.
- Allow venipuncture site to completely air dry after cleaning with alcohol. Alcohol damages cell walls.
- Place the needle correctly in the vein. Partial occlusion by the inner wall of the vein exerts shear force on the cells.
- Pull the plunger of syringe gently when drawing blood. Pulling too quickly exerts excess pressure and shear the cell walls.
- Pushing hard on the syringe plunger while transferring blood to a tube exerts a destructive level of pressure
- Avoid drawing from catheters and lines involve shear forces and turbulence cause hemolysis.
- Fill tubes to correct volume. Under-filling of tubes containing anticoagulant results in a higher than recommended concentration of the additive, which promotes hemolysis.
- Mix additives with the specimens by inverting tubes gently. Vigorous mixing or shaking can break the cells.
- Protect the specimens during transport. Exposure to inappropriate temperatures and significant jarring will cause hemolysis in transit.



Phiebotomy Essentials



Strasinger Di Lorenzo



Wolters Kluwer

Ruth E. McCall Cathee M. Tankersley

NOTES SECOND

Phlebotomy

POCKET GUIDE TO BLOOD COLLECTION

- Illustrated, step-by-step procedures
- Tube additive guide
- Best sites for blood collection
- Write-on, wipe-off



CLOTTED SAMPLES

EFFECTS OF CLOTTED SAMPLES:

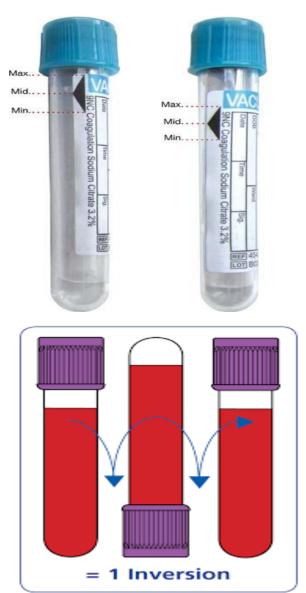
- Clots may alter composition of the specimens, producing inaccurate results.
- Clots can block the systems inside the analyzer, leading to analyzer breakdown and downtime → delay in turnaround time of test results and patient care.

CAUSES OF CLOTTED SAMPLES

- Delay of placing blood in tubes with additives e.g. leaving blood in a syringe too long before transferring to the tube.
- Improper mixing of blood with anti-coagulants in tubes or syringes (for blood gases).
- Over-filled tubes, leaving too little or no air-space that will enable proper mixing with anti-coagulants.

PREVENTION OF CLOTTED SAMPLES

- Transfer blood into the appropriate blood tubes immediately after collection.
- Fill the tubes with the recommended sample volume (as indicated on the tube).
- Gently invert the tubes to properly mix the blood with the anticoagulants (depends on type of tubes, usually 8-10 times).
- For blood gases, ensure the syringe is free of air bubbles and roll the syringe between your palms.



- Blood sample from a 60-year-old male taken in the outpatient setting revealed the following:
 - K⁺ 10.3 mmol/L
 - \circ Na⁺ and Cl⁻ normal
 - Total calcium 0.63 mmol/L
 - ALP 8 IU/L
 - Mg²⁺ 0.3 mmol/L
- Sample is not haemolysed. Patient is well.
- Cause?

○ K₂-EDTA contamination

 Important to know the ORDER OF DRAW to avoid cross-contamination of additives between blood tubes.



Order of Draw	Tube Stopper Colour	Tube Inversion	Rationale for the Collection Order
Blood culture (sterile collections)		8-10X	Minimise microbial contamination
Coagulation tubes	Light blue	4 X	The first additive tube in order because all other additives affect coagulation tests
Glass plain tubes	Red	Nil	Prevent contamination by other additives in other tubes. Plain glass tube should be drawn before the plastic serum tube with SST.
Clot activator, silicon coated (plastic) tubes	Red	5 X	Filled after coagulation tests because silica particles in the plastic tubes activate clotting and affect coagulation tests (carry-over of silica into
SST clot activator, gel separator tubes	Gold	5 X	subsequent tubes can be overridden by anticoagulant in them)
Heparin tubes	Green	8 X	Causes the least interference in tests other than coagulation tests
EDTA tubes	Lavender	8 X	Responsible for more carry-over problems than any additive. It elevated Na ⁺ and K ⁺ levels, chelates and decreases Ca ⁺⁺ and Fe levels, elevates PT and APTT results.
Oxalate / fluoride tubes	Gray	8 X	Oxalate is used after haematology tube (lavender stopper) because oxalate interferes in enzyme reaction, damages cell membranes and causes abnormal RBC morphology.

DELAYED SAMPLE PROCESSING

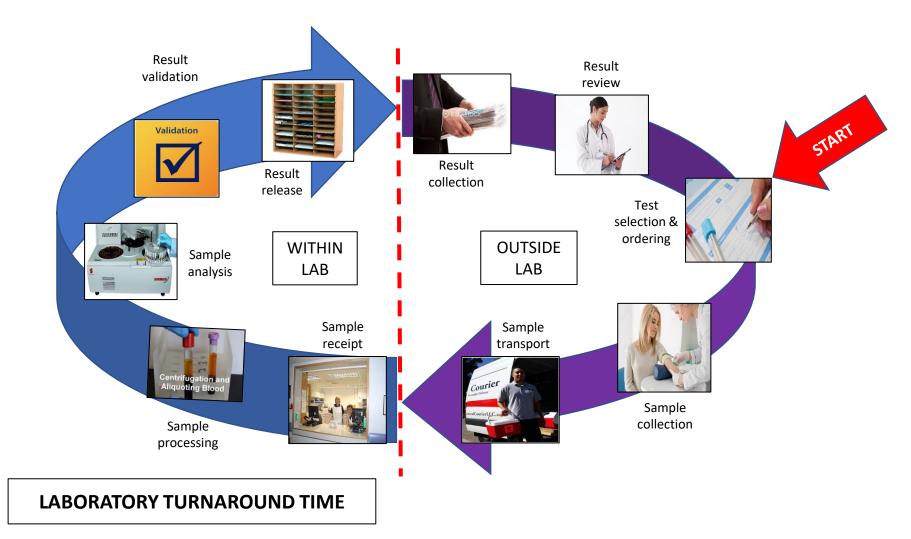
- The Clinical and Laboratory Standards Institute (CLSI) recommends that serum samples are separated within 2 hours of collection for most analytes.
- Some analytes are significantly affected by delayed separation because of:
 - Prolonged contact of plasma/serum with cells and leakage of intracellular constituents e.g. potassium, inorganic phosphorus, magnesium, LDH.
 - Glycolysis in cells which consumes **glucose** and produces **lactate**.
 - Degradation of **proteins** and **peptides** by blood enzymes.
- Urine samples stored at room temperature for > 2 hours will have:
 - Bacterial overgrowth
 - Lysis of RBCs, WBCs and casts
- Specimens need to be sent to the laboratory within the recommended time interval to ensure validity of test results.

LABORATORY TURNAROUND TIME (TAT)

= Total time taken from sample arrival at the lab to the release of validated result.

Type of request	TAT
Blood gases	45 minutes
hsTroponin T Urgent requests	1 hour
Inpatient but non-urgent requests	4 hours
Outpatient requests	5 working days
Special tests (run in batches) e.g. HbA1c, endocrine tests	5 working days
Outsourced tests	Depends on referral lab

LABORATORY TESTING PROCESS



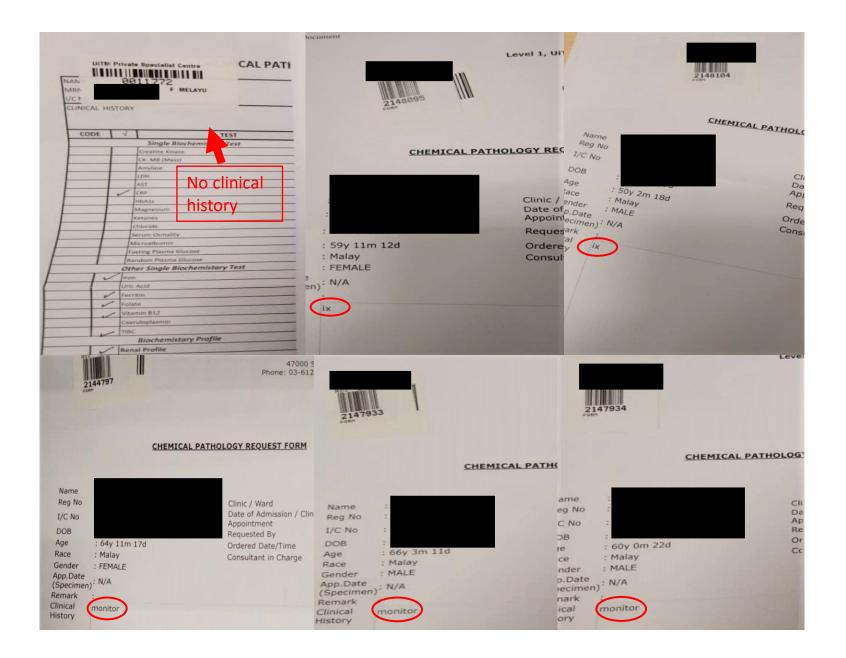
RESPONSIBLE REQUESTING



- Write relevant meaningful clinical history that may assist in interpretation of results.
- Know the indication when consulting the Pathologist for urgent / rare / expensive tests.
- Accept responsibility when notified about critical values (including from outpatients).
- Complete the required forms (e.g. PER PAT) when requesting for outsourced tests.



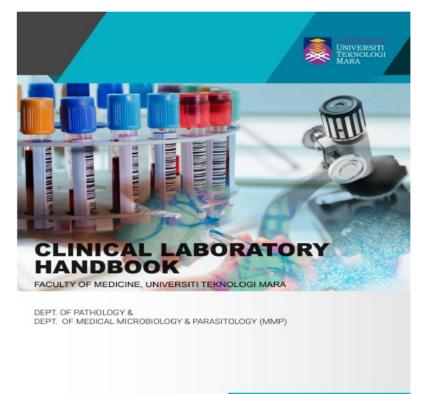
- Abuse urgent requests e.g. urgent request for health screening.
- Order tests without indication e.g. every single hormone or tumour marker on the list.
- Harass the lab staff to report results verbally. The policy of the lab is **NO verbal reporting** except for critical values.





- Add-on tests can be requested for samples that has been sent to the lab, provided:
 - ✓ adequate sample volume remains after the initial tests have been completed
 - stability of the analyte(s) requested are still acceptable.
- Please check with the lab staff first before ordering add-on tests via UniMEDS. A new request form should be sent to the lab for the add-on tests.
- For dynamic function tests, please inform the lab at least 1 day before performing the test and sending samples.

For further information, please refer to



1st EDITION

Downloadable from: https://pathologyuitm.weebly.com/

STANDARDS